Develoment of an Optimized PEI-based Transfection Reagent for the Production of Clinical Grade Viral Vectors: PEIpro®

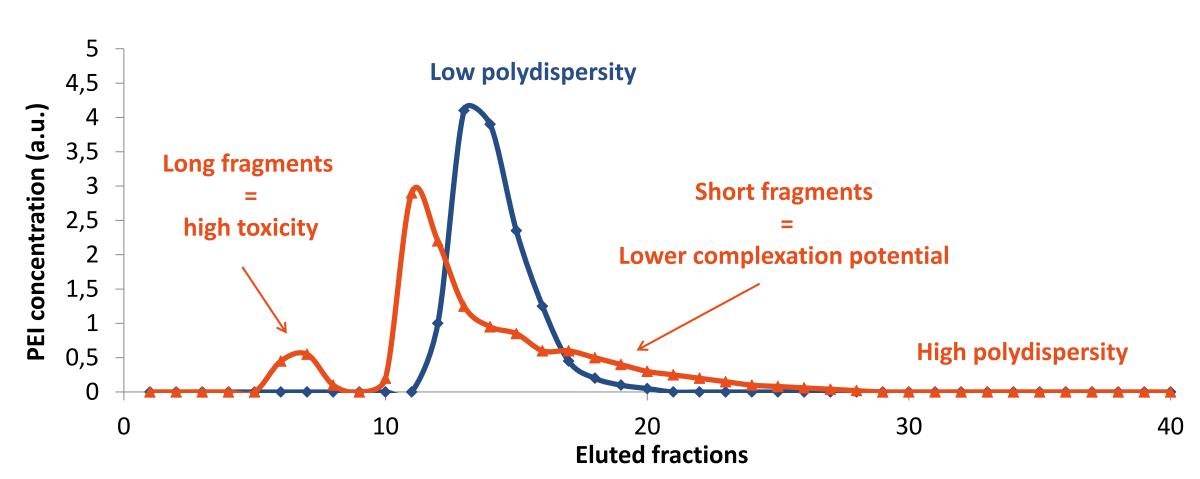


Géraldine Guérin-Peyrou, Mathieu Porte, Julien Depollier, Jonathan Havard, Patrick Erbacher Polyplus-transfection, Bioparc, 850 Boulevard S. Brant, 67400 Illkirch, France

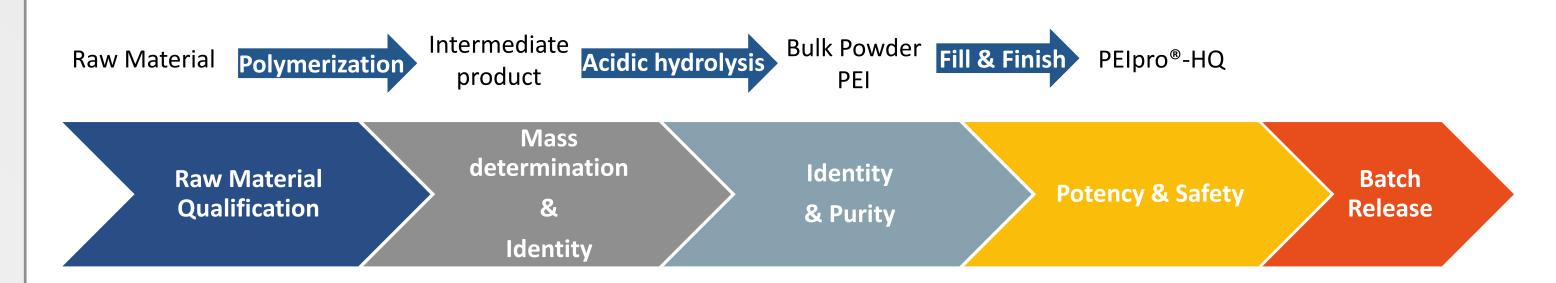
Abstract

Gene- and cell therapy-based medicine are experiencing resurgence due to the introduction of "next generation" transfer viral vectors, which have demonstrated improved safety and efficacy. Adeno Associated Virus (AAV) and Lentivirus are very commonly used in therapeutics and often produced using PEI-mediated transient transfection in HEK-293 or HEK-293T cells. PEIpro® and its high-quality counterpart PEIpro®-HQ are particularly well suited for high titer virus production. Both reagents are manufactured under a well-controled and reproducible process and are free of components of animal-origin. Here we show that they have been selected for their high transfection efficiency using low DNA amount. Moreover, they undergo stringent quality controls, making PEIpro® and PEIpro®-HQ the unique PEI-based transfection reagents suitable for use in process development and in cGMP biomanufacturing, respectively.

Optimized PEI for R&D to Clinical-Grade Virus Production



Optimization process of PEI polymer chemistry. Whereas long polymer fragments lead to cell toxicity and short fragments lead to lower complexation potential (in red), optimized PEI size with a low polydispersity index decreases toxicity, while increasing complexation potential (in blue) and reproducibility in transfection.



Manufacturing process of PElpro® and PElpro®-HQ reagents. The linear structure of PElpro® and the manufacturing process developed by Polyplus-transfection® ensure a high, stable and reproducible amount of protonable amines available for transfection while providing a fully deacylated molecule and an extremely low polymer chain length variation.

| | | PEIpro [®] | PEIpro®-HQ |
|---|--|---------------------|------------|
| Characteristics | | | |
| 1 mg/ml of Linear PEI in water | | ٧ | ٧ |
| Fully synthetic | | ٧ | ٧ |
| Manufacturing Process | | Identical | |
| Quality Controls | | | |
| | Molecular Weight of intermediate product | ٧ | ٧ |
| | Side chain content | | ٧ |
| Identity | Assay | | ٧ |
| | Color, Clarity | | ٧ |
| | рН | | ٧ |
| Purity | Impurity profile | | ٧ |
| | Endotoxin assay | ٧ | ٧ |
| Safety | Sterility test | ٧ | ٧ |
| | Mycoplasma | ٧ | v |
| Potency | Activity test | ٧ | V |
| Documentation | | | |
| Certificate of Analysis | | ٧ | ٧ |
| Certificate of Origin (confirming the absence of components of animal origin) | | ٧ | ٧ |
| Detailed Batch Production Documentation to include in an IND or IMPD | | | ٧ |

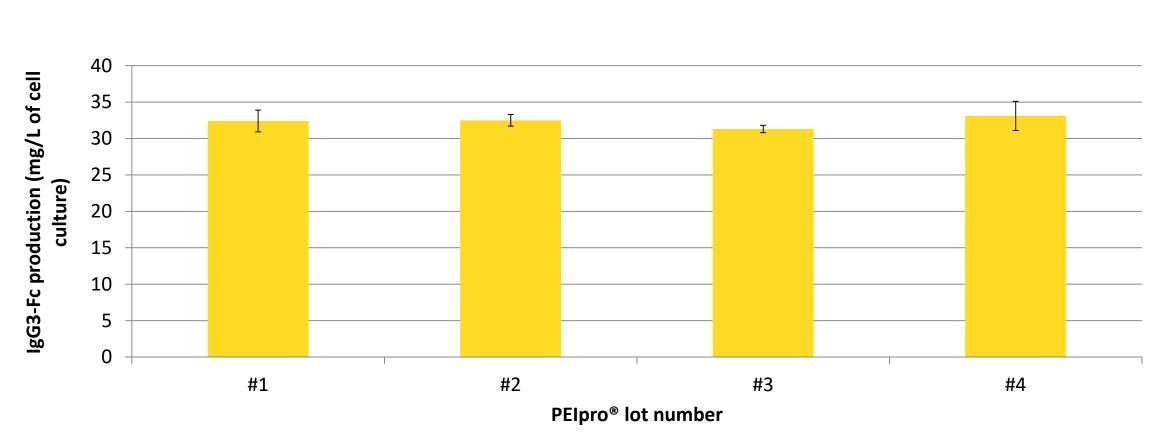
PEIpro®-HQ is a highly qualified grade of PEIpro® reagent. The quality of PEIpro® and PEIpro®-HQ are continuously assessed during the manufacturing process with suitable control testing. In comparison to PEIpro®, a more extensive number of quality controls are performed on both the bulk material and the formulated product of PEIpro®-HQ to assess **identity, purity, safety, and potency**.

Compatible with Various Production Culture Systems

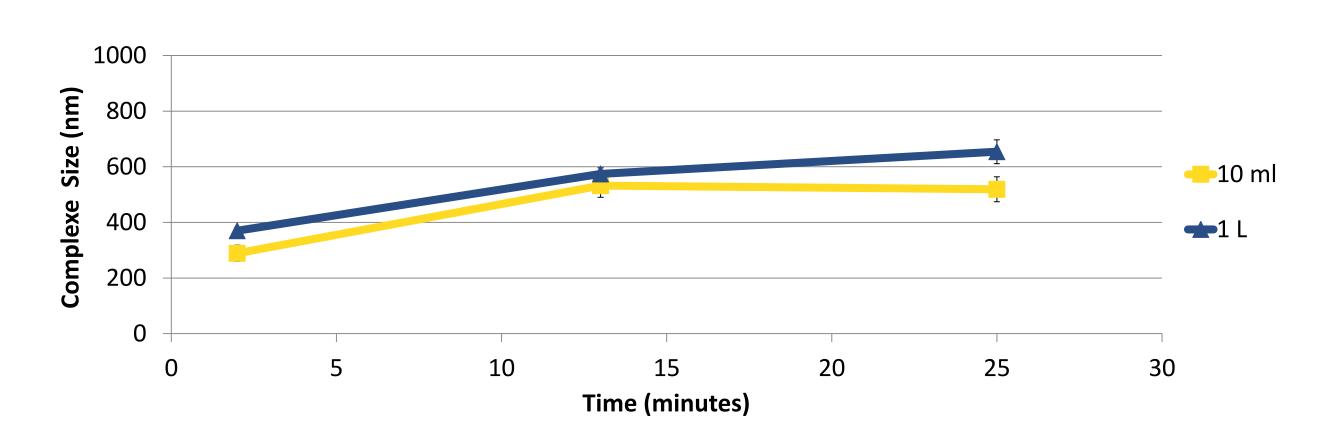
| Culture medium | Productivity using PEIpro® |
|---|----------------------------|
| DMEM (Gibco®) | +++ |
| IMDM (Gibco®) | +++ |
| Ham's F12 (Gibco®) | +++ |
| BalanCD® HEK293 (Irvine Scientific®) | +++ |
| FreeStyle™ 293 (Gibco®) | +++ |
| FreeStyle™ F17 (Gibco®) | +++ |
| HyClone™ HyCell™ TransFx™-H (GE Healthcare™) | +++ |
| Pro293™ (Lonza®) | ++ |
| CD 293 (Gibco®) | - |

PEIpro® in different media. PEIpro® and PEIpro® -HQ are compatible with several serum-containing media and commercially available synthetic media for virus production in both adherent and suspension cells HEK-293 cells.

Reproducibility & Scalability

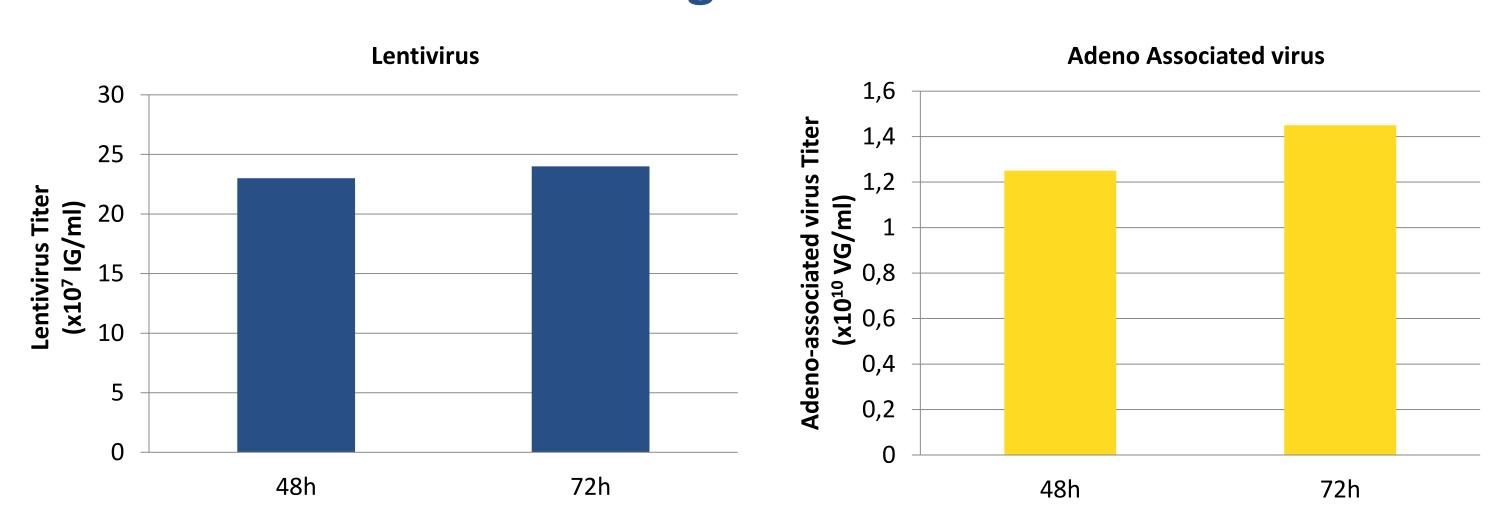


Excellent lot-to-lot protein yield reproducibility using PElpro[®]. Suspension HEK-293 cells were seeded at 1×10⁶ cells/mL in serum-free medium and transfected with PElpro[®] following the standard protocol. IgG3-Fc production was assayed 48 h after transfection using protein G affinity quantification (HPLC).



DNA-PElpro® complex size is identical, independently from the volume of transfection mix preparation. Complexes were prepared with a DNA concentration of 0.01 mg/mL of complex volume at a DNA:Reagent ratio of 1:4, either in 10 mL or 1 L. The size of the complexes was then measured at different time susing the Zetasizer Nanometer ZS (Malvern Instrument, Malvern, UK).

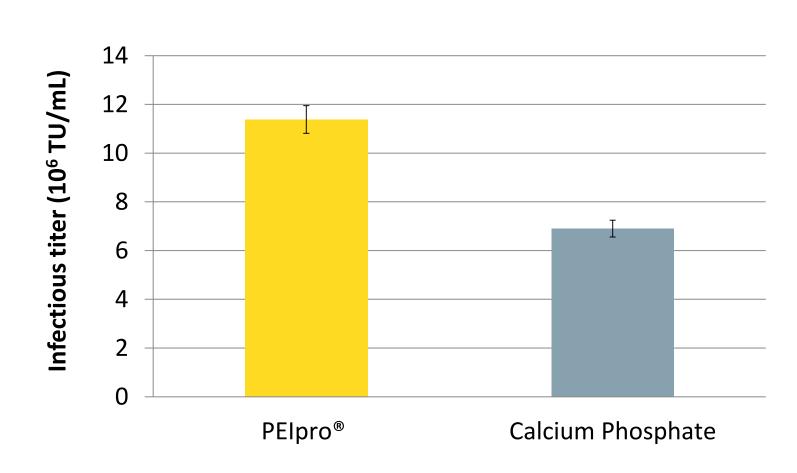
Gold standard for High Virus Production Yields



Lentivirus and Adeno Associated Virus (AAV) production in HEK-293T and HEK-293 cells grown in suspension in BalanCD® HEK-293 (Irvine Scientific®). HEK-293T (lentivirus) and HEK-293 (AAV) cells were thawed directly into each medium and passaged every 3 to 4 days before going into a 2 L benchtop bioreactor. Cells were seeded and cultured for 3 days before being transfected by PEIpro® (Polyplus-transfection®). For transfection, four plasmids were used for lentivirus and three plasmids were used for AAV. Lentiviral and AAV titer were measured 48 and 72 hours post transfection (Data kindly provided by Genethon).

| Cell culture system | Vector | Cells | Titer |
|---------------------------------|------------|-------------------------------|--|
| CS10® / CF10® | AAV | Adherent HEK-293, HEK-293T | 10 ¹¹ -10 ¹³ VG / mL |
| Fixed-bed bioreactor (iCELLIS®) | AAV | Adherent HEK-293T | 10^{14} - 10^{16} VG / mL |
| Shaker Flask | AAV | Suspension HEK-293, HEK-293T | 10 ⁹ -10 ¹⁰ VP / ml |
| Bioreactor | AAV | Suspension HEK-293, HEK-293T | 0.8 - 1.5×10^9 - $10^{10} \text{VG} / \text{mL}$ |
| 10 cm dish/75 cm2 | Lentivirus | Adherent HEK-293, HEK-293T | 1-2 x10 ⁸ TU / mL |
| HYPERflask®/HYPERstack® | Lentivirus | Adherent HEK-293, HEK-293T | 1-2 x10 ⁸ TU / mL |
| Shaker Flask | Lentivirus | Suspension HEK-293F, HEK-293T | 2x 10 ⁷ -10 ¹⁰ VP / mL |
| Bioreactor | Lentivirus | Suspension HEK-293, HEK-293T | 10 ⁷ IG / mL |

PElpro® is the reagent of choice for virus production runs in most cell culture systems in both adherent and suspension cells. Irrespective of the cell culture system and production scale, PElpro® and PElpro®-HQ have led to viral vector yields superior to 10⁷ IG/mL and 10⁹ VG/mL, for lentiviruses and AAV respectively.



PElpro® gives higher virus titers than Calcium Phosphate. Lentiviruses were produced in adherent HEK-293 cells grown in serum-free culture medium, using 15 μ g DNA and 30 μ L PElpro® per 75 cm² flask. Viral titers were determined by flow cytometry of supernatants 48 h after transfection.

Conclusion

Advantages of PElpro® and its higher quality grade PElpro®-HQ:

- A PEI optimized for transfection, suitable for virus production (and for protein production)
- Synthetic animal free reagent manufactured according to a well-established process
- Robust product with a great lot-to-lot reproducibility and a long shelf life
- Ideal for process development up to large-scale therapeutic viral vector production
- PEI of highest quality available with extra Quality Controls (identity, potency, safety and purity) and supplied with extra Quality extra GMP processes